UMBRACULUMIN-B, AN UNUSUAL 3-HYDROXYBUTYRIC ACID ESTER FROM THE OPISTHOBRANCH MOLLUSC UMBRACULUM MEDITERRANEUM

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Abstract: The structure and stereochemistry of Umbraculumin-B, a metabolite isolated from the skin extracts of the opisthobranch molluse Umbraculum mediterraneum, have been established by spectroscopic and chemical methods.

We have recently reported¹ the isolation and structure of two ichthyotoxic diacylglycerols, umbraculumin-A and -C, which could account for the chemical defense of the opisthobranch molluse *Umbraculum mediterraneum*. We whish to report now the structure of a third unrelated compound, named umbraculumin-B (1), accompanying the above compounds in the skin extracts of *U*. *mediterraneum* but inactive in the mosquito fish bioassay².

Fractions containing umbraculumin-B were obtained from the SiO_2 column chromatography also affording the other umbraculumins and were further purified by preparative TLC (benzene-diethyl ether; 7:3) where 1 is slightly more polar than umbraculumin-C; 4 mg of 1 were obtained from the skin extracts of the single specimen also used¹ for the isolation of umbraculumin-A and -C.

HREIMS showed that umbraculumin-B possesses the same molecular formula $C_{21}H_{32}O_5$ as umbraculumin-A (found M⁺ m/z 364.2228, calculated 364.2241) and was accompanied by a small amount of a compound m/z 366, formulated as the 2,3-dihydroderivative of 1 by careful integration of the appropriate ¹H-NMR signal (δ 5.45). The presence of four double bonds in 1 was inferred by a combination of UV and NMR data. The UV spectrum (*n*-hexane) has the characteristic shape of a conjugated triene chromophore with maxima at 279 (ϵ 28,000), 268 (ϵ 36,000) and 259 (ϵ 26,000) nm. The NMR proton signals for the central part of the triene moiety (from C-6 to C-9) were overlapped, even at 500 MHz, giving rise to a complex signal between 6.03 and 6.12 δ , while H-5 and H-10 were sufficiently distinct (δ 5.65 and 5.60, respectively) to allow assignments by decoupling experiments on irradiation of the appropriate adiacent methylene protons (see table). The large J values for the H-5/H-6 and H-10/H-9 couplings secured the *E* geometry for these double bonds. Although the ¹³C-NMR spectrum did not allow the assignment of the individual resonances from C-6 to C-9, the chemical shift values of these carbons (130.9; 131.5) indicated that the 7,8 double bond should have the *E* geometry, since consistently lower values are expected in a triene in which the central double bond has a *Z* geometry³.

H-2 and H-3 were also overlapped at δ 5.45 and coupled to the vinyl methyl at δ 1.65; this moiety was previously found also in umbraculumin-A and $-C^1$ and , similarly, the *E* geometry of the 2,3 double bond has been derived from the ¹³C chemical shift of the vinyl methyl⁴. Since irradiation of the C-4 methylene affects both the C-1,C-5 and the C-2,C-3 protons, the proton sequence from C-1 to C-10 was fully established; additional decoupling experiments also gave the C-10/C-13 proton sequence. The ¹H - ¹H COSY spectrum confirmed the above results.

TABLE.	NMR data	oſ	umbraculumin-B	(1) ^a
Position	δ ¹ Η		δ 130	:
(n	ultiplicity:	J.	Hz)	

	(
1	1.65 (m)	18.0
2	(5.45 (m)	126.4 ^b
3		128.9 ^b
4	2.78 (m)	35.8
5	5.65 (m;J ₅₋₆ 14.2)	133.0
6	r	,
7	6.03-6.12 (m)	131.5 (2C)
8		130.9 (2C)
9	l	l
10	5.60 (m;J ₁₀₋₉ 15.0)	132.4
11	2.15 (q; 7.4)	29.2
12	1.72 (m)	28.4
13	4.10 (t; 6.6)	64.4
14		171.8°
15	2.60 (ddd;15.3, 7.8, 4.	7) 40.8
16	5.35 (m)	67.7
17	1.33 (d; 5.9)	20.0
18		170.5°
19	2.47 (ddd;16.5, 8.6, 2.9	9) 43.5
20	4.20 (m)	64.4
21	1.20 (d; 6.1)	22.5

^aCDCl₃; Bruker WM 500. ^{b,c}Values with identical superscripts may be interchanged.



The ¹³C-NMR spectrum showed the presence of two ester carbonyls at δ 170.5 and 171.8 and of three sp³ carbons linked to oxygen, one CH₂ (δ 64.4) and two CH (δ 64.4 and 67.7) by DEPT sequence. These functions were readily accomodated in two β -hydroxybutyric acid residues linked to each other,one of them esterifying the tridecatetraenol residue, by observing the C-15/C-17 and C-19/C-21 proton sequences both by ¹H-¹H COSY and by proton decoupling experiments. In particular the C-15 and C-19 methylenes resonated in the ¹H-NMR spectrum as well resolved AB parts (8 lines each) of two distinct ABX systems, the X

parts (H-16 and H-20) being coupled to the two methyl doublets at δ 1.33 and 1.20 respectively. A ¹H-¹³C HETCOR spectrum allowed the assignment of the individual ¹³C resonances as reported in the table.

The absolute configuration at C-20 was determined to be R as follows. Umbraculumin-B (3 mg) dissolved in toluene (0.5 ml) was reacted (80°C; 16h) with an excess (20 μ l) of (R)(-)-1-(1-naphtyl)ethylisocyanate. After elimination of the solvent the crude product was dissolved in 0.5 ml of 1N H₂SO₄/CH₃OH and allowed to react (r.t.; 4 days). The solution was neutralized with solid NaHCO₃ and the product purified by preparative TLC (CH₂Cl₂/EtOAc, 95:5) to afford the uretane 2. By using the above chiral isocyanate the 1-(1-naphtyl)ethyl uretanes of (R,S)- and (S)-3-hydroxybutyric acid methyl ester were prepared by conventional procedures. Their ¹H-NMR spectra (CDCl₃) differ mainly in the chemical shift value of the carboxymethyl group which resonates at δ 3.70 in the R,R diastereomer and δ 3.58 in the R,S diastereomer. The uretane 2 derived from umbraculumin-B displays a δ 3.70 value for the carboxymethyl group and hence the R stereochemistry was inferred for the 3-hydroxybutyric acyl residue. The stereochemistry at C-16 is also tentatively designed as R since it is obvious to expect that the two 3-hydroxybutyric acyl residues may have the same absolute stereochemistry.

Whereas umbraculumin-A and -C are very toxic to fish at 10 and 0.1 μ g/ml respectively¹, umbraculumin-B is inactive at 10 μ g/ml.

REFERENCES

1. G. Cimino, A. Crispino, A. Spinella and G. Sodano, Tetr.Lett. 29, 3613, 1988.

2. L. Gunthorpe and A.M. Cameron, Mar.Biol. 94, 39, 1987.

3. F.W. Wehrli and T. Nishida, Progress in the Chemistry of Organic Natural Products 36, p.128, 1979.

4. J.B. Sthothers, "Carbon-13 NMR Spectroscopy", Academic Press, New York, 1972, p.81.

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